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Identification study some virulence factors of invasive mold infections isolated from patients undergoing chemotherapy in Tikrit teaching Hospital

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ABSTRACT

Sixty two (62) samples were collected from out and inpatients admitted to Tikrit teaching hospital in Tikrit city. these specimens were collected from sputum of immunocopromised patients of both sexes and different ages.

19(30.6%) isolates from patients sputum, were infections more *A. fumigatus* (12.9%), *A. terreus* (6.5%), and *P. sp* (3.3%) while were (1.6%) of *A. niger*, *A. flavus*, *F. solani*, *P. chrysogenum* and *A. alternata*. The isolated were identified according morphological and Cultural characteristic.

It emerges from the study that fungal isolates produced various virulence factors since, hemolysin, protease and phospolypase production were produced in *A. fumigatus* (62.5%) hemolysin and phosholipase production, and (87.5%) protease production, while *A. terreus* protease and phosholypase production were (50%) and (25%) of hemolysin production, while *A. flavus*, *P. chrysogenum* and *A. alternata* isolates were no produced protease. *P. chrysogenum* were produce only hemolysin (100%). All isolates of *A. niger*, *F. solani* and *A. alternata* were phospolypase production (100%).

INTRODUCTION

Fungi are ubiquitous in the environment. Fungi that are part of the normal flora, or that are wide spread in the environment we are constantly exposed to fungal spores (conidia), and we inhale on a regular basis and are normally harmless to immunocompetent individuals. Some groups of fungi are pathogenic to humans and require control measures. Human fungal pathogens belong to four main groups, namely zygomcetes, ascomycetes, deuteromycetes, and basidiomycetes (Chakrabarti, 2005; Reedy *et al.*, 2007).

Invasive fungal infection (IFI) is increasingly being recognized as a significant cause of morbidity and mortality in immunosuppressed patients (Garcia-Vidal *et al.*, 2013).

Immunocompromised patients who are less capable individuals of battling infections because of an immune response that is not properly functioning (Christoph, 1991).

Invasive infections due to filamentous fungi, such as *Aspergillus* spp., *penicillium* sp., *Fusarium* spp. and *Alternaria* sp. cause significant morbidity and mortality in immunocompromised patients with hematological malignancies, recipients of hematopoietic stem cell transplants and those with chronic granulomatous disease (Antachopoulos *et al.*, 2012).

Fungi are able to cause a disease and to overwhelm the host defense systems because of possessing several genes and proteins associated with their pathogenicity, called virulence factors (Tomee & Kauffman, 2000). Many of the putative fungal virulence factors have developed naturally during organism evolution and originally acted as a defense against unfavorable environmental conditions, and then, in this way, many of them became important as virulence factors facilitating infection (Casadevall & Pirofski, 2000), this study aimed to:

- Isolation and identification of mold infection from immunocompromised patients.
- Screening the ability of mold isolates to produce, protease, hemolysin and phospholypase as a virulence factors.

MATERIAL AND METHOD

1- Samples Collection: Sputum and blood samples were collected from 62 patients undergoing chemotherapy within age range "11-83 years old", were during chemotherapy for different types of cancer attending Tikrit Teaching Hospital from January 2013 to Augusto 2013 were enrolled in this study.

2- Direct Examination: Specimen were placed on a microscopic slide, with the few drops of 10% KOH, a cover slip added and warmed over a small flame just before boiling. The slide was examined under the low power and high dry objectives to detect fungi and their septet hyphae (Emmons *et al.*, 1977).

3- Samples Culturing: Sputum samples were cultured on sabouraud dextrose agar (SDA)

supplemented with 0.04 mg/ml chloramphenicol to inhibit the growth of bacteria, then incubated at 28°C and 37°C and examined for 10 days (Midgley *et al.*, 1997).

4- Criteria Of Growth Identification: All isolates were identified to the species level on the basis of macromorphological and micromorphological characteristics using SDA, and Scotch tape preparation.

Identification of the growth depends on the following:

1- Colony characteristics (Color, Consistency and Topography).

2- Colony Reverses (Color, Significant pigment).

3-Microscopic Morphology (Microconidia and Macroconidia: their Size, Shape, Arrangement, and Hyphal Structures). (Atlas *et al.*, 1995).

4. Haemolysin Activity test: Determination of haemolysin activity was evaluated with a blood plate assay according Manns *et al.*, 1994

5. Protease Production test: The protease production was determined according to Aoki 1990 using a test medium consisting of SDA plates containing bovine serum albumin (BSA).

6. Phospholipase Production: Phospholipase activity assays were performed according to (Price *et al.*, 1982).

RESULTS AND DISCUSSIO

Results fungal isolates in patients undergoing chemotherapy at tikrit teaching hospital were 19(30.6%) positive from total examined patients.

Sputum samples results showed that *A. fumigatus* were most frequently isolated (12.9%), followed by *P. sp.* (9.6%), *A. terreus* (9.%), and *Penicillium* sp. (3.3%) all each for them , *A. niger*, *A. flavus* and *P. chrysogenium* *F. solani* and *A. alternata* (1.6%) were frequently less isolated recoding , as illustrated in Table (1).

Table 1: Number and percentage of identified fungal isolates from sputum culture samples.

Fungal isolates	Number of isolates	%
<i>fumigatus</i>	8	12.9
<i>niger</i>	1	1.6
<i>terreus</i>	4	6.5
<i>flavus</i>	1	1.6
<i>P. chrysogenum</i>	1	1.6
<i>F. solani</i>	1	1.6
<i>alternata</i>	1	1.6
<i>P. sp</i>	2	3.3
No growth	19	63.4
Total	62	100

Our results for sputum culture showed the *Aspergillus* spp. were more 14(22.5%) out of 62 specimen.

There were many studies which have been conducted on opportunistic fungi in immunocompromised patients. One of these studies was Manahil *et al.*, (2011) in mosul who was isolated Four species of the genus *Aspergillus* were detected from sputum in immunocompromised patients. The total isolates were 9(4.5%), 4 of them were *A. flavus*, 2 *A. fumigatus*, 2 *A. niger*, and 1 *A. terreus*. *Penicillium* spp. 3(1.5%), were *Fusarium* spp. 1(0.5%).

As reported Al-Khalidi and Faraj (2012) (56.66%) isolates of sputum were that isolated included *Aspergillus niger* and *A. fumigatus* 3 out of 34(8.82%) for each one, *A. flavus*, *Penicillium* spp. 2 out of 34 (5.88%) for each one and *A. alternate* 1 out of 34 (2.94%).

In other study were 26 (27%) of *C. albicans* and were 29 (30%) non *albicans* spp., *Aspergillus* spp. 6.25% were *A. fumigatus* and *A. niger* each of them, were 1% of *A. flavus* and followed by *Penicillium* spp. in 6 (6.25%) including two *P. marneffei* isolates (Bharathi and Usha., 2011).

The reason for these variations in all studies may be due to sample size, environment factors, nutrition requirements and virulence factors of this fungi (Cooke *et al.*, 2009), and methods used for isolation.

It is not easy to determine the pathogenic role of fungal isolates from the respiratory tract, to differentiate between infections, colonization and contamination (Paradowski, 1997). However, the

prevalence and prognosis of pulmonary fungal infection is difficult to be evaluated since diagnosis was seldom confirmed (Chen *et al.*, 2001).

An important factor contributing to the increasing incidence of infection by fungi that have not been previously described to be pathogenic, is the rise in numbers of immunocompromised patients who are susceptible hosts for the most uncommon microbial agents (Disalvo., 2005).

Mentioned that Invasive *Aspergillus* is an opportunistic fungus, the immunocompromised patients are more susceptible for its colonization. Whether infection (aspergillosis) could happen, it is a matter of balance between the immunity of the person versus the pathogenicity of the organism. Denning and Coworker *et al.* (2003).

2. Identification of molds isolates:

The isolates identified performed a morphological characterization using the coloration on SDA as indicating that each species has a specific color and For descriptions of species, and additional information see Samson and Pitt (1990), de Hoog *et al.* (2000) and Ellis *et al.* (2007).

A: Identification of *A. alternate*:

A. alternate was able to grow and form colonies on SDA media in a moderate growth rate. The colony was felt, the color at the first seem off-white to be to then turning to dark brown and changed to grey-brownish, reversed side of the colonies appeared pale yellow to tan Figures (4-5A,B,C).

Microscopic examination as shown in Figure (1) multicelled, septate and irregularly branched. Conidiophores arised singly or in clusters, usually were long or short. They were pale olivaceous to, conidia were brown with a short, cylindrical beak, and form long and profusely branched chains (ten or more conidia). Similar results were reported by Wellman (1949), Ghosh (1998) and Nagrale *et al.* (2012).

B: Identification of *Aspergillus* species:

The isolates showed different apparent morphological characteristics, mainly in the color of the culture medium and For descriptions of species showed Table (2) and the Figures (2 to 5) shows image which *Aspergillus species* isolates during this study The present findings are in conformity with the result of earlier workers Nelson *et al.* (1983), Samson and Pitt (1990), de Hoog *et al.*, (2000) and Ellis *et al.*, (2007).

Table 2: Morphology and Microscope feature of *Aspergillus* species.

Species	Surface	Reverse	Conidiophore	Phialides	Vesicle
<i>A. flavus</i>	Yellow-green	Coldish to red brown	Colourless, rough	Uniseriate or biseriate	Round, radiate head
<i>A. fumigatus</i>	Blue-green to gray	White to tan	Short, smooth, colourless or greenish	Uniseriate	Round, columnar head
<i>A. niger</i>	Black	White to yellow	Long, smooth, colourless or brown	Biseriate	Round, radiate head
<i>A. terreus</i>	Cinnamon to brown	White to brown	Short, smooth, colourless	Biseriate	Round, compactly columnar head

C: Identification of *F. solani*:

Morphotype produced aerial to abundant cottony mycelium with pale white to cream, brown-greenish to white-greenish aerial mycelium, and the pigmentations were pale brown to yellowish brown with a dark brown zonation.

Microscope examinations showed Figure (6) Microconidia are usually abundant, cylindrical to oval, one-to two-celled and formed from long lateral macroconidial septation from 3 to 5 from short multi-branched.

D: Identification of *P. chrysogenum*:

The colonies on SDA Medially grown. The colonies were velvety and sulcate, with blue-green in colour. The reverse side of the colonies was yellow, as shown in Figure (7).

Microscopically, penicilli were terverticillate and the conidia were spherical to elliptical in shape. Conidia were smooth and had a green colour reflection. The results agree with other studies such as, Justen *et al.*, (1998) and we found ability isolate to growth

at 37°C that was in a agreement with Rafi and Rahman (2002) but do not agree with Florey *et al.* (1949) and Singh *et al.* (1991).

1- Screening of some fungi isolates for virulent factors.

In order to select the efficient isolate for some virulent factors production, screening of fungal isolates had been achieved on specific media.

A: Screening of some fungi isolates for protease production.

In this study, the extracellular protease production of *A. fumigtus* (87.5%), (50%) of *A. turreus* and (100%) of *A. niger* while none *A. favus* isolate produce protease , which is in agreement with (Theeb *et al.*, 2013) who stated that hemolysin is produced by (95.1%) from 41 *A. fumigatus* isolates. Also, In Another study connected by Aboul-Nasr *et al.* (2013) who found 20 (80%) of *A. falvus*, 16(62%) of *A. fumigatus*, 19(79%) and 17(65%) of *fusarium solani*, as shown in Table (3).

Table 3: test protease Production by fungal isolates.

Fungal isolates	Number isolates	Protease	
		No	%
<i>A. fumigatus</i>	8	7	87.5
<i>A. terreus</i>	4	2	50
<i>A. niger</i>	1	1	100
<i>A. flavus</i>	1	0	0
<i>F. solani</i>	1	1	100
<i>A. alternata</i>	1	0	0
<i>P. chrysogenum</i>	1	0	0

In this study, the extracellular protease production of *A. fumigatus* (87.5%), (50%) of *A. terreus* and (100%) of *A. niger* while none *A. flavus* isolate produce protease, which is in agreement with (Theeb *et al.*, 2013) who stated that hemolysin is produced by (95.1%) from 41 *A. fumigatus* isolates. Also, In Another study connected by Aboul-Nasr *et al.* (2013) who found 20(80%) of *A. flavus*, 16(62%) of *A. fumigatus*, 19(79%) and 17(65%) of *fusarium solani*.

Salyers and Witt (1994) reported that microbial cells secrete hydrolytic enzymes that destroy the constituents of host cell membranes leading to membrane dysfunction, physical disruption as well as aid in the invasion of host tissues. Proteolytic degradation of lung tissues

has been suggested as one of the key events involved in the physiopathology of *A. fumigatus* (Kothary *et al.*, 1984).

B: Screening of fungi isolates for hemolysin production:

Our results indicated that (62.5%) of *A. fumigatus*, (25%) of *A. terreus*, then *A. niger*, *A. flavus* and *P. chrysogenum* were (100%) for each of them and (50%) of *F. solani*., as shown in Table 4 and Figure (8). Which are In agreement with the study of Aboul-Nasr *et al.* (2013) who found 20(95%) of *A. flavus*, 16(75%) of *A. fumigatus*, 16(68.5%) of *A. niger* and 17(41.2%) of *F. solani*. However in other study it has been found that (73.1%) *P. chrysogenum* were produces Hemolysin (Rossoni *et al.*, 2013).

Table 4: hemolysin Production some by some fungal isolates.

Fungal isolates	Number isolates	Hemolysis	
		No	%
<i>A. fumigatus</i>	8	5	62.5
<i>A. terreus</i>	4	1	25
<i>A. niger</i>	1	1	100
<i>A. flavus</i>	1	1	100
<i>F. solani</i>	2	1	50
<i>A. alternata</i>	1	0	0
<i>p. chrysogenum</i>	1	1	100

These differences are possible to be due to the environmental conditions, the source of isolate and the detection method that was used in the detection of the enzyme.

Hemolysin lyses RBCs by creating pores or holes in red blood cell membranes resulting in the release of iron that promotes microbial growth (Bullen, 1981; Almeida *et al.*, 2009).

C: Screening of fungal isolates for phospholipase production:

Results showed that (62.5%) *A. fumigatus*, (50%) of *A. terreus* and (100%) of *A. niger*, *F. solani* and *A. alternata* of each for them isolates were positive for phospholipase production, as shown in Table (5).

Table 5: Production of phospholipase by fungal isolates.

Fungal isolates	Number isolates	Phospholipase	
		No	%
<i>A.fumigatus</i>	8	5	62.5
<i>A.terreus</i>	4	2	50
<i>A.niger</i>	1	1	100
<i>A.flavus</i>	1	0	0
<i>F.solani</i>	1	1	100
<i>A.alternata</i>	1	1	100
<i>p. chrysogenum</i>	1	0	0

Our results were relatively agreed with the results of Birinci *et al.* (2014) was detected of phospholipase activity in 30(93.3%) of *A. fumigatus*, 4(25%) and None of nine *A. flavus* isolates exhibited phospholipase activity.

Phospholipases may enhance pathogenesis of invasive fungi by hydrolysing phospholipids in the membrane lipid bilayer of epithelial and endothelial cells. Phospholipids in mammalian cell membranes and in lung surfactant are preferred substrates for secreted phospholipase B from *C. albicans* and hence are a potential site of membrane attack (Chen *et al.*, 2000).

Reports from different countries show that there are more different phospholipase activities in different regions (Zarei *et al.*, 2010) Borst and Fluit (2003) believed that virulence factors could be associated with geographical region and infection type.

Probably variable levels of phospholipase activity in our study are related to source of fungal isolates (Oksuz *et al.* 2007), Borst and Fluit (2003) also found different phospholipase activity between samples originated from urine, blood and wound.

REFERENCE

- Aboul-Nasr, M.d, Aboul-Nasr B. Abdel-Naser A. ohri2 and Amer E. M. (2013): Enzymatic and toxigenic ability of opportunistic fungi contaminating intensive care units and operation rooms at Assiut University Hospitals, Springer Plus., 2:347-349.
- Al-Khalidi A. A., M. J. and M. K. Faraj (2012): Isolation and identification of fungi associated with chronic respiratory infections in human and bovine. College of Veterinary Medicine\ University of Baghdad Al-Anbar J. Vet. Sci., 5(2):1999-6527.
- Almeida RS, Wilson D. and Hube B. (2009): *Candida albicans* iron acquisition within the host. FEMS Yeast Res. 9(7):1000-12.
- Antachopoulos, C., Katragkou, A Roilides, E. (2012): Immunotherapy Against Invasive Mold Infections Immunotherapy., 4(1):107-120.
- Aoki S, Ito-Kuwa S, Nakamura Y. and Masuhara T. (1990): Comparative pathogenicity of wild-type strains and respiratory mutants of *Candida albicans* in mice. Zol. Bakt., 273: 332-43.
- Atlas R. M., Williams J. F., Huntington M. K. (1995). *Legionella* contamination of dental-unit waters. *Appl. Environ. Microbiol.* 61 1208–1213.
- Bharathi M. and A. Usha Rani (2011). Pathogenic fungal isolates in sputum of HIV positive patients ISSN 2141-2359 .2011 Academic Journals.
- Birinci A., Bilgin K., Çayci Y. T. (2014): Investigation of Acid Proteinase and Phospholipase Activity as Virulence Factors in Clinical Aspergillus spp. Isolates. Mikrobiyol Bul, 48(3): 491-494.
- Borst A, Fluit AC. (2003): High levels of hydrolytic enzymes secreted by *Candida albicans* isolates involved in respiratory Infections. J. Med Microbiol. 52: 971-4.

- Bullen JJ. (1981): The significance of iron in infection. *Rev Infect Dis.*, 3: 1127-1138.
- Casadevall A. and Pirofski LA. (2001): Host-pathogen interactions: the attributes of virulence. *J Infect Dis* 184:337–344.
- Chakrabarti A (2005) Microbiology of systemic fungal infection. *J Postgrad Med* 51(Suppl1): Chemother 56:5–11.
- Chen KY, Ko SC, Hsueh PR. (2001): Pulmonary fungal infections. Emphasis on microbiology spectra, patient outcome, and prognosis factors. *Chest*. 120: 177-184.
- Chen, S.C.A., Wright, L.C., Golding, J.C. and Sorrell, T.C. (2000): Purification and characterization of secretory phospholipase B and lysophospholipase / transacylase from a virulent strain of the pathogenic fungus *Cryptococcus neoformans*. *J. Biochem.* 347:431-439.
- Cooke NS, Feighery C, Armstrong DK, Walsh M, Dempsey S. (2009) Cutaneous *Fusarium solani* infection in childhood acute lymphoblastic leukaemia. *Clin Exp Dermatol* 34:e117–e119.
- De Hoog, G.S., Guarro, J., Gené, J., Figueras, M. J. (2000): Atlas of clinical fungi Centraalbureau voor Schimmecultures, 1126ff.
- Denning DW, Riniotis K, Dobrashian R, (2003): Chronic cavitary and fibrosing pulmonary and pleural aspergillosis: Case series, proposed nomenclature change and review. *Clin Infect Dis*. 37(3): 265.
- Disalvo A. (2005): Opportunistic mycoses. *Mycology*-chapt, seven.
- Ellis, B.J. (2007): Pubertal timing, Heritability, and Environmentality: A Critical Look. Paper presented at the biennial meeting of the Society for Research in Child Development, Boston, MA.
- Emmons, C. W.; Binford, C. H.; Utz, J. P. and Kowchong, K. J. (1977): *Medical Mycology*. 3rd Ed. Lea and Febiger, Philadelphia.
- Frolich, S., Entzeroth, R., and Michael Wallach (2011): Comparison of Protective Immune Responses to Apicomplexan Parasites. Vol. (2012): Article ID 852591, 11 pages.
- Garcia-Vidal, Carolina; Viasus, Diego; Carratalà, Jordi, (2013): Pathogenesis of invasive fungal infections. *current infections diseases opinion infect*, 26(3): 213-293.
- Justen, P., G.C. Paul, A.W. Nienow and C.R. Thomas, (1998): Dependence of *Penicillium chrysogenum* growth, morphology, vacuolation and productivity in fed-batch fermentations on impeller type and agitation intensity. *Biotechnol. Bioeng.*, 59: 762–75.
- Kothary, R. K., Burgess, E. A. And Candido, E. P. M. (1984): The heat shock phenomenon in cultured cells of rainbow trout hsp70 mRNA synthesis and turnover. *Biochim. biophys. Acta* 783:137–143.
- Manahil M. Yehia, Zainalabideen A. Abdulla (2011): Opportunistic fungi in lower respiratory tract infection among immunocompromised and immunocompetent patients College of Medicine, University of Mosul. (Ann. Coll. Med. Mosul 2012; 38 (1): 59-67.
- Manns JM, Mosser DM, Buckley HR. (1994): Production of a Hemolytic Factor by *Candida albicans*. *Infect Immun*. 62:5154-6.
- Midgley, G.; Clayton, Y. M. and Hay, P. R. J. (1997): *Diagnosis in color medical mycology*. 1st Ed., Mosby-Wolf.
- Nagrle, D. T., Gaikwad, A. P., Goswami, S. and Sharma, L. (2012): Fungicidal management of *Alternaria alternata* (Fr.) Keissler causing blight of gerbera (*Gerbera jamesonii* H. Bolus ex J.D. Hook). *J. Appl. & Nat. Sci.*, 4 (2): 220-227.

- Nelson, P.E.; Toussoun, T. A. and Marasas, W.F.O. (1983): *Fusarium species: An Illustrated Manual for Identification*. Pennsylvania State University Press, University Park. 193pp.
- Oksuz S, Sahin I, Yildirim M, (2007): Phospholipase and proteinase activities in different *Candida* species isolated from anatomically distinct sites of healthy adults. *Jpn J Infect Dis.* 60: 280-3.
- Paradowski LJ. (1997): Saprophytic fungal infections and lung transplantation – revisited. *J Heart lung Transpl.*, 16: 524-531.
- Price MF, Wilkinson ID, Gentry LO. (1982): Plate method for detection of phospholipase activity in *Candida albicans*. *Sabouraudia.* 20: 7-14.
- Rafi, M. and Rahman S. U. (2002): Isolation and Identification of Indigenous *Penicillium chrysogenum* Series / *Int. J. Agri. Biol.*, Vol. 4, No. 4.
- Reedy JL, Bastidas RJ, Heitman J (2007): The virulence of human pathogenic fungi: notes from the Richardson MD (2005): Changing pattern and trends in systemic fungal infections. *J. Antimicrob S16–S20* South of France. *Cell Host Microbe* 2:77–83.
- Rossoni RD, Barbosa JO, Vilela SFG, Jorge AOC, Junqueira JC. (2013): Comparison of the hemolytic activity between *C. albicans* and non-*albicans* *Candida* species *Braz Oral Res.*, Sao Paulo, 27(6):484-9 485.
- Salyers A, Witt D (1994): Virulence factors that damage the host. *Bacterial pathogenesis: a molecular approach.* ASM Press, Washington, D.C, pp. 47–62.
- Samson, R.A. and Pitt, J.I. eds. (1990): *Modern Concepts in Penicillium and Aspergillus Classification.* Plenum Press: New York and London, 478 pp.
- Singh, B.; Bobogare, A.; Cox-Singh, J.; Snounou, G.; Abdullah, M. S. and Abdul Rahman, H. (1999): A Genus - And Species- Specific Nested Polymerase Chain Reaction Malaria Detection Assay For Epidemiological Studies. *The American Society of Tropical Medicine and Hygiene.* 60(4): 687–692.
- Wellman, F.L. (1949): Successful spray control of *Alternaria* blight of petunias grown for seed in Costa Rica. *Pl. Dis. Repr.*, 33(2): 69-72.
- Zarei Mahmoudabadi A, Zarrin M, Miry S. (2010): Phospholipase activity of *Candida albicans* isolated from vagina and urine samples. *Jundishapur J Microbiol.* 3(4): 169-73.



Fig. 1: Morphology of *A.alternata* grown on SDA at 37°C A: the Colony morphology after 10 days top view B: Colony morphology after 10 days reversed view C: Microscopic morphology (40x).

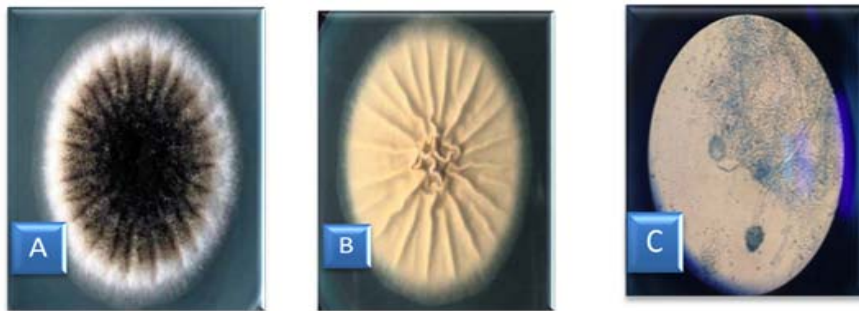


Fig. 2: Culture morphology of *A. niger* grown on SDA after 5 days at 37°C A: Top view B: Reversed view C: Microscopic morphology of *A. niger* stained by gram stain (40x)



Fig. 3: Morphology of *A. terreus* grown on SDA at 37°C A: The colony morphology after 5 days Top view B: The colony morphology after 5 days Reversed view C: Microscope morphology of *A. terreus* staining by LPCB stain (40x).

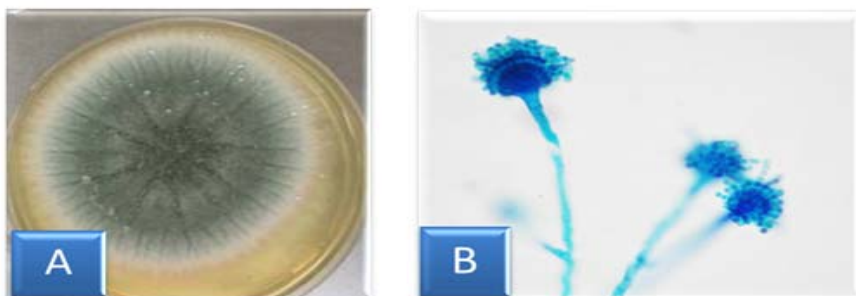


Fig. 4: A: the colony morphology of *A. fumigatus* grown on SDA at 37°C B: microscope feature of *A. fumigatus* staining by LPCB stain (40x).

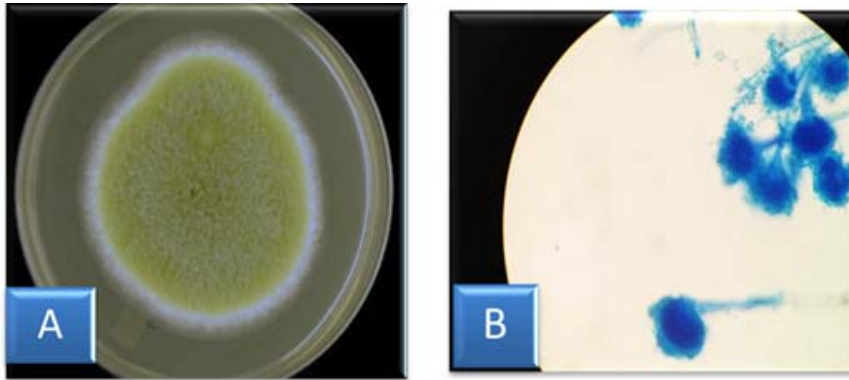


Fig. 5: A: the colony morphology of *A. flavus* grown on SDA at 37°C B: Microscope morphology of mycelium staining by LPCB stain (40x).

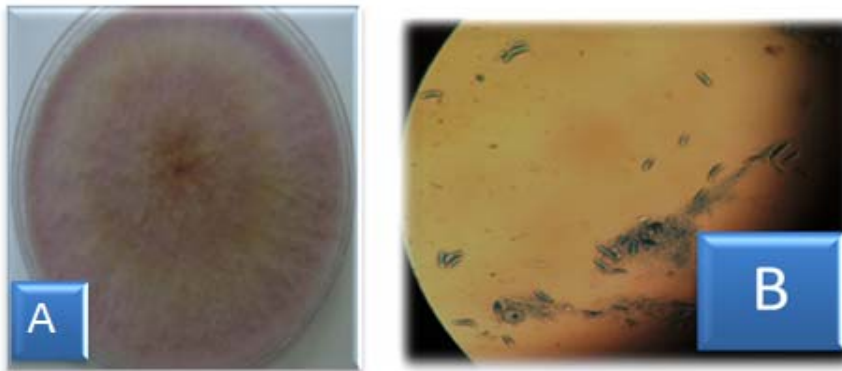


Fig. 6: A: Culture morphology of *F. solani* grown on SDA after 7 days B: Microscope morphology of *F. solani* grown on SDA at 37°C on SDA staining by LPCT stain (40x).



Fig. 7: Culture morphology of *P. chrysogenum* grown on SDA after 7 days.

ARABIC SUMMARY

دراسة تحري عن بعض عوامل الضراوة من الاعفان الغازية المعزولة في المرضى الخاضعين للعلاج الكيماوي في مستشفى تكريت التعليمي.

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 ٢- كلية الصيدلة / جامعة تكريت
 ٣- علوم الحياة / كلية العلوم / جامعة تكريت

تم جمع ٦٢ عينة قشع من المرضى الراقدين والمراجعين لمستشفى تكريت التعليمي من كلا الجنسين ومن مختلف الاعمار ضمن هذه الدراسة.

كانت نسبة الإصابة 19(30.6%) عزل من القشع ، كانت اعلى نسبة اصابات هي *A. fumigatus* بنسبه (12.9%) وكانت *A. terreus* بنسبه (% (6.5), *P. sp* بنسبه (% (٣.٣) . بينما كانت العزلات من *A. niger* و *A. flavus* , *F. solan* و *P. chrysogenum* و *A. alternata* بنسبه (% (١.٦) لكل منها حيث تم تشخيص العزلات وفقا لخواصها الشكلية والمزرعية.

تم دراسة عوامل الضراوة متنوعه للفطريات المعزولة مثل انتاج protease و hemolysin و phospholipase كانت الانتاج في *A. fumigatus* بنسبه (% (62.8) لانتاج كل من hemolysin و phospholipase وبنسبة (% (87.5) منتجة لل hemolysin ، بينما *A. terreus* كانت منتجة لل phospholipase و protease في بنسبه (% (٥٠) و انتاجها hemolysin كانت بنسبه (% (٢٥) ، بينما عزلات *A. flavus* و *P. chrysogenum* و *A. alternata* كانت غير منتجة لل protease بينما كل عزلات *A. niger* و *F. solani* و *A. alternata* كانت منتجة phospholipase بنسبه (% (١٠٠).